



Early View

Research letter

Reduced immunogenicity of the mRNA vaccine BNT162b2 in patients with IPF

Theodoros Karampitsakos, Ourania Papaioannou, Ilias Dimeas, Panagiota Tsiri, Vasilina Sotiropoulou, Ioannis Tomos, Ilias C Papanikolaou, Matthaios Katsaras, Paraskevi Kirgou, Zoe Daniil, Konstantinos I Gourgoulianis, Fotios Sampsonas, Effrosyni Manali, Spyridon Papiris, Demosthenes Bouros, Argyris Tzouvelekis

Please cite this article as: Karampitsakos T, Papaioannou O, Dimeas I, *et al.* Reduced immunogenicity of the mRNA vaccine BNT162b2 in patients with IPF. *ERJ Open Res* 2022; in press (<https://doi.org/10.1183/23120541.00082-2022>).

This manuscript has recently been accepted for publication in the *ERJ Open Research*. It is published here in its accepted form prior to copyediting and typesetting by our production team. After these production processes are complete and the authors have approved the resulting proofs, the article will move to the latest issue of the ERJOR online.

Copyright ©The authors 2022. This version is distributed under the terms of the Creative Commons Attribution Non-Commercial Licence 4.0. For commercial reproduction rights and permissions contact permissions@ersnet.org

Reduced immunogenicity of the mRNA vaccine BNT162b2 in patients with IPF

Theodoros Karampitsakos¹, Ourania Papaioannou¹, Ilias Dimeas², Panagiota Tsiri¹,
Vasilina Sotiropoulou¹, Ioannis Tomos³, Ilias C Papanikolaou⁴, Matthaios Katsaras¹,
Paraskevi Kirgou², Zoe Daniil², Konstantinos I Gourgoulianis², Fotios Sampsonas¹,
Effrosyni Manali³, Spyridon Papiris³, Demosthenes Bouros⁵, Argyrios Tzouvelekis¹

¹ Department of Respiratory Medicine, University Hospital of Patras, Greece

² Department of Respiratory Medicine, Medical School, University of Thessaly, Larissa,
Greece

³ 2nd Pulmonary Medicine Department, "ATTIKON" University Hospital, Athens Medical
School, National and Kapodistrian University of Athens, Athens, Greece

⁴ Respiratory Medicine Department, "Corfu General Hospital", Corfu, Greece

⁵ First Academic Department of Pneumonology, Hospital for Thoracic Diseases, "SOTIRIA",
Medical School, National and Kapodistrian University of Athens, Athens, Greece

Correspondence to:

Argyrios Tzouvelekis MD, MSc, PhD

Associate Professor of Respiratory Medicine

Head Department of Respiratory Medicine

University of Patras, Greece

argyrios.tzouvelekis@fleming.gr

Take home message: Patients with IPF did not mount appreciable antispikes antibody responses to two doses of SARS-CoV-2 mRNA vaccine compared to general population. National authorities should prioritize patients with IPF for booster doses.

To the editor:

The emergence and spread of 2019 coronavirus disease (COVID-19) are causing a growing global public health crisis. Despite advances in treatment, vaccination remains the best way to contain the pandemic¹. Vaccines are currently available by means of conditional marketing approval, full approval and emergency use authorization pathways². Evidence suggest that immunocompromised individuals including solid organ transplants recipients and patients under immunosuppressive treatment may have increased mortality from SARS-CoV-2 infection despite double dose messenger RNA (mRNA) vaccine regimens³. This is partially attributed to blunted immune responses to vaccination since only 38-54% of kidney and liver transplant recipients developed detectable SARS-CoV-2 antibodies following the second dose of mRNA vaccines^{3,4}.

Patients with Idiopathic Pulmonary Fibrosis (IPF) present with disrupted cellular and humoral immune responses⁵. In view of previous data during the first waves of the pandemic indicating increased risk of death from COVID-19 in unvaccinated patients with fibrotic ILDs⁶, we aimed to determine humoral responses to two doses of SARS-CoV2 BNT162b2 mRNA vaccine in patients with IPF and compare them to those seen in the general population.

Therefore, we conducted a multicenter, prospective study between 22/7/2021 and 31/10/2021 including patients who received a multidisciplinary diagnosis of IPF in 5 Interstitial Lung Disease centers in Greece and aged-matched controls. Prospective data collection and analysis was approved by the Institutional Review Board and the Local Ethics Committee (protocol number 18482/21-7-21). IPF was diagnosed based on ATS/ERS 2018 Guidelines⁷.

We compared anti-SARS-CoV-2 antibodies three months after the second dose of the mRNA vaccine BNT162b2 in three subgroups: 1) patients with IPF receiving antifibrotics, 2) patients with IPF under no treatment, 3) aged-matched controls. Patients receiving corticosteroids or steroid sparing agents and patients with malignancies were excluded from the analysis. Subjects with history of prior SARS-CoV-2 infection were also excluded. The recruitment period was between 22/7/2021 and 31/10/2021. All measurements were performed using the authorized Abbott SARS-CoV-2 IgG assay. Positivity cut-off threshold for this assay was 50 AU/ml.

With regards to summary statistics, categorical data were presented as absolute numbers and relative frequencies. Continuous data were demonstrated as mean± standard deviation (SD) or medians with 95% Confidence Interval (95% CI) based on Kolmogorov-Smirnov test for normality. Kruskal-Wallis test was used to detect differences in the aforementioned three subgroups. P-values < 0.05 were considered statistically significant.

Sixty-seven (n=67) subjects were included in the analysis (patients with IPF receiving antifibrotics: 32, patients with IPF under no treatment: 10, controls: 25). Groups were age and gender balanced [median age- IPF/treatment: 72.5 (95% CI: 69.9 to

75.0) vs IPF/no treatment: 73.0 (95% CI: 67.9 to 80.5) vs controls: 69.0 (95% CI: 65.5 to 74.2), $p=0.15$), gender- IPF/treatment: males 78.1% ($n=25/32$), IPF/no treatment: males 80% ($n=8/10$), controls: males 76% ($n=19/25$)]. Median Forced Vital Capacity% predicted and diffusing capacity of the lung for carbon monoxide %predicted were 81.0 (95% CI: 68.8 to 89.4) and 47.0 (95% CI: 40.9 to 57.4), 106.9 (95% CI: 66.1 to 121.1) and 60.7 (95% CI: 50.7 to 85.1), as well as 91.5 (95% CI: 87.0 to 97.8) and 87.0 (95% CI: 80.0 to 97.0) in the IPF/treatment, IPF/no treatment and control groups, respectively. The majority of subjects were current or ex-smokers in all groups [81.3% ($n=26/32$), 80% ($n=8/10$) and 76% ($n=19/25$), respectively]. Arterial hypertension was the most common comorbidity among all groups [43.8% ($n=14/32$), 40% ($n=4/10$) and 40% ($n=10/25$), respectively], while proportions of all comorbidities including obesity were balanced among groups. Both groups of patients with IPF whether receiving or not antifibrotic compounds exhibited similarly reduced levels of anti-SARS-CoV-2 antibodies after two doses of the mRNA vaccine BNT162b2 compared to general population [666.1, (95%: 540.1 to 900.0) vs 579.5 (95% CI: 232.4 to 4054.2) vs 2118.6 (95% CI: 1248.3 to 4035.5), $p=0.002$] (**Figure 1**). The prevalence of anti-SARS-CoV-2 antibodies above the suggested cut-off threshold of 1000 AU/ml was 21.9%, 40% and 72% in the IPF/treatment, IPF/no treatment and control groups, respectively.

To the best of our knowledge, this is the first study showing that patients with IPF did not mount appreciable antispikes antibody responses to two doses of SARS-CoV-2 mRNA vaccine compared to general population. Importantly, impaired immunogenicity was observed irrespective of anti-fibrotic treatment further adding to the knowledge that current anti-fibrotics do not cause further

immunosuppression but rather confer protection to disease acute exacerbations including those infection-mediated^{8,9}. This report is of interest given that impaired immune response following vaccination in most diseases has not been attributed to the disease per se but to the compounds used to treat the disease. Our findings are in line with mechanistic data showing blunted cellular and humoral immune responses in patients with IPF potentially contributing to fibrosis development and progression. In particular, patients with IPF and poor disease outcomes present with T-cell exhaustion as indicated by down-regulation of T-cell co-stimulatory markers. Impressively, the same high-risk genomic profile was also associated with increased mortality in patients with COVID-19¹⁰, indicating that similar aberrant immune responses predispose to severe SARS-CoV-2 pneumonitis and fibrotic ILD. The relationship between B cells and IPF remains largely speculative, given the conflicting results of previous studies. On the one hand, increased detection of CD20+ B cells has been reported in lungs of patients with IPF¹¹. Detection of B lymphocyte stimulator, denominated B-cell-activating factor (BAFF), is enriched in the peripheral blood and pulmonary parenchyma of patients with IPF, while neutralization or genetic ablation of BAFF attenuated pulmonary fibrosis¹². On the other hand, data derived from other animal models demonstrated that B cells may suppress fibrotic response¹³.

Limitations of our study include a moderate sample size that may lack generalizability, lack of serial measurements and underpowered arm of patients with IPF under no treatment. However, treatment-naive patients with IPF represent only a small minority in the era of antifibrotics and thus our sample size of non-treated IPF patients is representative of the everyday clinical practice. Moreover, we had not

measured anti-SARS-CoV2 antibody levels prior to vaccination; yet, we excluded subjects with history of SARS-CoV-2 infection. Another limitation is that we used Abbott test which quantifies IgG directed against an epitope of the spike protein, but we did not investigate specific immune responses such as specific T cell responses. We did not also record the exact time of the day that vaccination was performed to adjust for the effect of circadian rhythm. Finally, there is a small age difference between patients with IPF and controls; nonetheless, this is not statistically significant.

Our findings are of important value and should prompt national authorities to prioritize those subgroups of patients for booster doses and stringent precautions at the level of other groups of immunosuppressed individuals considering that reduced levels of anti-SARS-CoV-2 protective antibodies as well as fibrotic lung diseases per se are independent risk factors for increased mortality following SARS-CoV-2 infection^{6,14}. Additional precautionary measures should include personal protective approaches such as protective facial masks and awareness of physical distancing, that played a beneficial role in the early phases of the pandemic¹⁵; yet, emergence of highly contagious variants of concerns mandate vaccination approaches to boost effectiveness and encourage fully vaccinated household contacts to offer optimal protection for at risk individuals.

References

1. Karampitsakos T, Malakounidou E, Papaioannou O, et al. Tocilizumab improves 28-day survival in hospitalized patients with severe COVID-19: an open label, prospective study. *Respiratory research*. Dec 22 2021;22(1):317.
2. Polack FP, Thomas SJ, Kitchin N, et al. Safety and Efficacy of the BNT162b2 mRNA Covid-19 Vaccine. *New England Journal of Medicine*. 2020.
3. Kamar N, Abravanel F, Marion O, Couat C, Izopet J, Del Bello A. Three Doses of an mRNA Covid-19 Vaccine in Solid-Organ Transplant Recipients. *New England Journal of Medicine*. 2021;385(7):661-662.
4. Boyarsky BJ, Werbel WA, Avery RK, et al. Antibody Response to 2-Dose SARS-CoV-2 mRNA Vaccine Series in Solid Organ Transplant Recipients. *Jama*. Jun 1 2021;325(21):2204-2206.
5. Idiopathic Pulmonary Fibrosis Clinical Research N, Raghu G, Anstrom KJ, King TE, Jr., Lasky JA, Martinez FJ. Prednisone, azathioprine, and N-acetylcysteine for pulmonary fibrosis. *The New England journal of medicine*. May 24 2012;366(21):1968-1977.
6. Drake TM, Docherty AB, Harrison EM, et al. Outcome of Hospitalization for COVID-19 in Patients with Interstitial Lung Disease. An International Multicenter Study. Dec 15 2020;202(12):1656-1665.
7. Raghu G. R-JM, Myers JL, Richeldi L, Ryerson CJ, Lederer DJ, Behr J, Cottin V, Danoff SK, Morell F, Flaherty KR, Wells A, Martinez FJ, Azuma A, Bice TJ, Bouros D, Brown KK, Collard HR, Duggal A, Galvin L, Inoue Y, Jenkins RG, Johkoh T, Kazerooni EA, Kitaichi M, Knight SL, Mansour G, Nicholson AG, Pipavath SNJ, Buendía-Roldán I, Selman M, Travis WD, Walsh S, Wilson KC. Diagnosis of Idiopathic Pulmonary Fibrosis. An Official ATS/ERS/JRS/ALAT Clinical Practice Guideline. *American journal of respiratory and critical care medicine*. Sep 1 2018;198(5):e44-e68.
8. Collard HR, Richeldi L, Kim DS, et al. Acute exacerbations in the INPULSIS trials of nintedanib in idiopathic pulmonary fibrosis. *The European respiratory journal*. May 2017;49(5).
9. Ley B, Swigris J, Day B-m, et al. Pirfenidone Reduces Respiratory-related Hospitalizations in Idiopathic Pulmonary Fibrosis. *American journal of respiratory and critical care medicine*. 2017;196(6):756-761.
10. Juan Guardela BM, Sun J, Zhang T, et al. 50-gene risk profiles in peripheral blood predict COVID-19 outcomes: A retrospective, multicenter cohort study. *EBioMedicine*. Jul 2021;69:103439.
11. Todd NW, Scheraga RG, Galvin JR, et al. Lymphocyte aggregates persist and accumulate in the lungs of patients with idiopathic pulmonary fibrosis. *Journal of inflammation research*. 2013;6:63-70.
12. François A, Gombault A, Villeret B, et al. B cell activating factor is central to bleomycin- and IL-17-mediated experimental pulmonary fibrosis. *Journal of autoimmunity*. Jan 2015;56:1-11.
13. Arras M, Louahed J, Simoen V, et al. B lymphocytes are critical for lung fibrosis control and prostaglandin E2 regulation in IL-9 transgenic mice. *American journal of respiratory cell and molecular biology*. May 2006;34(5):573-580.
14. Gilbert PB, Montefiori DC, McDermott A, et al. Immune Correlates Analysis of the mRNA-1273 COVID-19 Vaccine Efficacy Trial. *medRxiv*. 2021:2021.2008.2009.21261290.
15. Papiris SA, Bouros D, Markopoulou K, et al. Early COVID-19 Lockdown in Greece and IPF: A beneficial “impact” beyond any expectation. *European Respiratory Journal*. 2020:2003111.

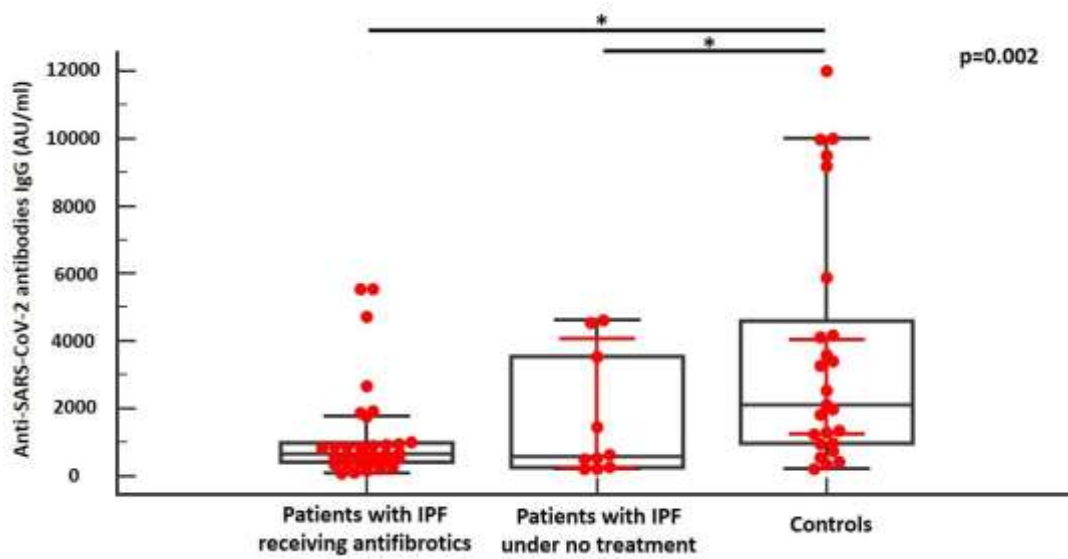


Figure 1. Both patients with IPF receiving antifibrotics and patients with IPF under no treatment had reduced levels of anti-SARS-CoV-2 antibodies after two doses of SARS-CoV-2 mRNA vaccine compared to general population (Kruskal-Wallis test, $p=0.002$).