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Early View

Invited review

ERS international Congress 2023: highlights from the Basic and Translational Sciences Assembly

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ERS international Congress 2023: highlights from the Basic and Translational Sciences Assembly

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Take home message

ERJOR: In case you missed the #ERS2023, this article highlights key messages of the @EuroRespSoc @3Assembly: Basic and Translational Sciences sessions that were discussed in Milan! @SaraOcana1 @EarlyCareerERS @ERSpublications

Abstract

In this article, early career members of the Assembly 3: Basic and Translational Sciences of the European Respiratory Society summarise the key messages discussed during six selected sessions that took place at the ERS congress 2023 in Milan, Italy. Aligned with the theme of the congress, the first topic covered is 'micro- and macro-environments and respiratory health', followed by a summary of the 'Scientific year in review' session. Next, recent advances in experimental methodologies and new technologies were highlighted within the 'tissue modelling and remodelling' session, as well as summary of the translational science session, 'what did you always want to know about omics analyses for clinical practice?', organised as part of the ERS Translational Science Working group's aims. Details on how the next-generation sequencing can be integrated with laboratory methods were provided in the 'lost in translation: new insights into cell-to-cell crosstalk in lung disease' session and a final summary of studies presented in the 'from the transcriptome landscape to innovative preclinical models in lung diseases' session linking the transcriptome landscape with innovative preclinical models was included in this review. The wide range of topics covered in the selected sessions and the high quality of the research discussed highlight the strength of the basic and translational science being presented at the international respiratory conference organised by the ERS.

Abbreviations

Alveolar type 2 (AT2) Alveolar type 1 (AT1) Aryl hydrocarbon receptor (AHR) Birt-Hogg-Dubé syndrome (BHD) Black carbon (BC) Bronchoalveolar lavage (BAL) Charcot-Leyden crystals (CLC) Chronic obstructive pulmonary disease (COPD) Cystic fibrosis (CF) Cystic fibrosis transmembrane conductance regulator (CFTR) Cytometry by time-of-flight (CyTOF) Digital spatial profiling (DSP) European Respiratory Society (ERS) Fibrotic hypersensitivity pneumonitis (fHP) Forced expiratory volume in 1 second (FEV₁) Forced vital capacity (FVC) Fractional exhaled nitric oxide (FeNO) Human antigen R (HuR) Human induced pluripotent stem cell (hiPSC) Human precision-cut lung slices (hPCLS) Idiopathic pulmonary fibrosis (IPF) Interstitial lung diseases (ILDs) IPSC-derived AT2 (iAT2) Isthmin-1 (ISM1) Keratin 17 (KRT17) Matrix metallopeptidase (MMP) Multiple Iterative Labeling by Antibody Neodeposition (MILAN) Mycobacterium tuberculosis (Mtb) Neutrophil trap (NET) Particulate matter $\leq 2.5 \,\mu m (PM_{2.5})$ Particulate matter $\leq 10 \ \mu m \ (PM_{10})$ Phosphatase and tensin homolog (PTEN) Post-COVID pulmonary fibrosis (PCPF) RNA sequencing (RNAseq) Single-cell RNA sequencing (scRNAseq) Single-nuclei RNA sequencing (snRNAseq) Surfactant protein C (SP-C) Surfactant protein C (SP-C) gene (SFTPC) Terminal airway-enriched secretory cells (TASCs) Tissue inhibitor of metalloproteinase (TIMP) Transient receptor potential vanilloid 4 (TRPV4) Type 2 helper T (Th2) cell Volatile organic compounds (VOCs)

The European Respiratory Society (ERS) International Congress 2023 took place in a hybrid form, hosting more than 20,000 participants, attending either in person (17,309 registrations) in Milan, Italy or participating online (3,215 registrations). The congress focused on tackling key areas of respiratory medicine: pollution, climate change and sustainable developments. As in previous years [1-3], there were numerous types of sessions including oral presentations, symposia, hot topics, poster presentations and year in review [4]. In this article, early career members of the ERS Assembly 3 [5] summarise some of the most relevant sessions describing the latest state of the art technologies, and sessions giving insights into the future direction of basic and translational respiratory science. Additional content can be accessed on the virtual platform (https://live.ersnet.org/home/ers/ers2023/en-GB).

Micro- and macro-environments and respiratory health - Oral presentation session

The respiratory system is closely linked with the environment. Various elements in the surroundings impact lung function, ranging from endotypes, the microbiome, and the microenvironment, to the macroenvironment, including indoor and outdoor air pollution and green spaces. This also encompasses the utilization of inhaler treatments. These factors interact with one another within a complex network, influencing respiratory health as assessed by spirometry measurements such as forced vital capacity (FVC) and forced expiratory volume in 1 second (FEV₁), fractional exhaled nitric oxide (FeNO), symptom burden, allergic rhinitis, chronic obstructive pulmonary disease (COPD), and the incidence of asthma (Fig. 1).

In this session, Dr Helena Backman (Luleå, Sweden) showed that COPD endotypes can be identified through different lung function trajectories with unique biomarker profiles. Three trajectories (T1: mean age 65 years, ever-smokers 72%, T2: mean age 58 years, ever-smokers 100%, T3: mean age 71 years, ever-smokers 78%) were described exhibiting combinations of different FEV₁, mean high sensitive C-reactive protein, matrix metallopeptidase (MMP)-9, and MMP-9/tissue inhibitor of metalloproteinase (TIMP)-1 ratio values [6, 7]. Miss Beatrice Cornu Hewitt (Utrecht, Netherlands) presented a study assessing the association between livestock-related emissions, (e.g., bacteria, antimicrobial resistance genes), and the oropharyngeal-acquired resistome (defined as an inherited set of genes used to resist infections) structure in COPD individuals versus healthy individuals. This study showed that the individuals' with COPD airway exhibited a higher resistome diversity, while *E.coli* was associated with significant differences in the oropharyngeal resistome of all individuals in the study [8-11]. Dr Randi Bertelsen (Bergen, Norway) explained the link between the exposure to indoor bacteria and lung function and inflammation in children. More specifically, higher microbial diversity is associated with better lung function (measured by FVC and FEV₁ z-scores) in males and increased inflammation (measured by FeNO) and lower lung function in females [12].

Dr Zhebin Yu (Stockholm, Sweden) next showed an association between air pollution exposure and long COVID symptoms. This study combined an estimation of particulate matter $\leq 2.5 \ \mu m (PM_{2.5}), \leq 10 \ \mu m (PM_{10})$, black carbon (BC), and nitrogen oxide levels with the evaluation of long COVID symptoms acquired via questionnaires from 753 participants. Exposure to PM_{2.5} was linked to long COVID, dyspnea, and altered smell/taste [13-15]. Dr Carlos Valencia-Hernandez (London, United Kingdom) presented the association of urban green spaces with lung function in ages from six to sixteen years in three European birth cohorts. Despite the high heterogeneity between the studies, the presence of green environment was linked to a small increase in FEV₁ and FVC values, although analysis of further cohorts is ongoing [16]. Dr Inês Paciência (Oulu, Finland) explained the role of exposure to air pollution as a modifier on the association between access and exposure to green spaces and development of allergic rhinitis. A study including 2568 participants demonstrated the beneficial role of green spaces, which is more important in cases of high air pollution exposure [17]. Miss Rina So (London, United Kingdom) explained the risk of COPD and asthma in relation to air pollution. This study investigated the Danish population and the annual mean levels of PM_{2.5}, NO₂, and BC. Higher exposure was related to higher asthma and COPD incidence defined by hospital contact [18-21].

Finally, Dr Joachim Heinrich (Munich, Germany) presented a study on long-term exposure to ambient ozone in 3014 adults from 17 centers in nine countries. Higher exposure was associated with faster lung functional decline estimated by spirometry [22, 23]. Dr Henning Kothe (Hamburg, Germany)

explained the impact of cooking methods on indoor air pollution and lung function in rural Rwanda. Indeed, replacing traditional cooking with improved cookstoves resulted in a reduction in IAP and an improvement in lung function [24]. Finally, Dr Joan B Soriano (Madrid, Spain) showed the estimated economic burden and carbon footprint in metric tons of CO₂ equivalent of the change of inhalers for non-clinical reasons and the consequent lack of adherence to treatment in Spain, producing a great economic and environmental cost [25].

Scientific year in review – Year in Review

In this Scientific year in review session, the speakers summarised the latest advances in translational respiratory science made by labs from across the world over the last year. Dr Rosa Faner (Barcelona, Spain) demonstrated the importance of gene-environment interactions in the pathogenesis of COPD. Being born severely pre-term (<28 weeks gestation age) was associated with a 7-fold increased risk of developing COPD by age 30-50 [26]. This links to another study of preterm children who underwent COPD polygenic risk scoring. Those who had the highest risk scores developed reduced FEV₁ at the age of 5, which shows that COPD-associated genes may play a role in preterm children developing obstructive airways disease [27].

Dr Faner also discussed the role of epigenetics in COPD. A study focussing on ethnically-diverse children living in low-income areas identified a genetic variant that was partly mediated by DNA methylation changes associated with smoking history – this variant was associated with reduced FEV₁ [28]. In addition, studies continue to show the importance of telomere shortening in the development of COPD [29]. This led to an insightful conversation amongst the panellists about the potential to screen for individuals at risk of COPD using telomere length and polygenic risk scores.

Dr Maor Sauler (Connecticut, United States) presented the latest research into alveolar defects in obstructive lung disease. He discussed that pro-inflammatory macrophages are associated with ferroptosis of alveolar type 2 (AT2) epithelial cells in lungs exposed to cigarette smoke [30]. In addition, recent data show that the loss of zinc transporter ZIP8 results in impaired AT2 cell function and subsequent lung fibrosis. Exogenous zinc then renewed the activity of AT2 cells, raising the potential of zinc as a therapeutic target in idiopathic pulmonary fibrosis (IPF) [31]. Previous studies have shown that transfection with specific miR-200 family members (including miR-200c-3p) restored transdifferentiation of AT2 cells obtained from people with IPF to alveolar type 1 (AT1) cells [32]. More recent research showed that this is through down-regulation of the endothelial Flt1 receptor [33]. Flt1 knock-out mice were protected from lung fibrosis upon exposure to bleomycin, and fibrosis was even reversed in these mice [33].

Dr Sauler showed research focusing on small airway disease in COPD. Single-cell RNA sequencing (RNAseq) (scRNAseq) identified a new cell type found in distal airways, termed 'terminal airwayenriched secretory cells' (TASCs), which secretes surfactant. There is loss of TASCs in the distal airways of end-stage COPD individuals, which may contribute to the loss of distal airways seen in COPD [34].

Dr Melanie Königshoff (Pennsylvania, United States) focused on anti-ageing targets in IPF (Fig. 2). Airway basal cells in IPF are reprogrammed to a keratin 17 (KRT17)^{high} and phosphatase and tensin homolog (PTEN)^{low} cell type. These cells contributed to fibrosis development when implanted into mouse lungs, changes that were attenuated by the Src kinase inhibitor saracatinib [35]. Saracatinib,

initially developed as an oncological treatment, reverses several fibrotic pathways – a trial of saracatinib in IPF is ongoing [36, 37].

Dysfunction of the endothelial transcription factor ERG occurs during aging, and was associated with increased systemic inflammation, vascular remodelling and impaired lung fibrosis recovery following bleomycin administration [38]. Lower levels of another endothelial transcription factor, FOXF1, were observed in endothelial cells obtained from people with IPF. FOXF1-deficient endothelial cells were associated with accelerated lung fibrosis and inflammation, and lung delivery of FOXF1 cDNA via nanoparticles attenuated lung fibrosis development in mice treated with bleomycin, showing the potential of this finding as a treatment strategy in IPF [39]. These results emphasise the importance of the lung endothelium in ageing and the pathogenesis of IPF.

Dr Wolfgang Kübler (Berlin, Germany) presented advances in the understanding of tissue barrier dysfunction in pathogen-associated respiratory failure. He described that the matrikine endostatin is increased in the lungs of ARDS individuals, including COVID-19-related ARDS, and this promoted thrombin-induced epithelial barrier dysfunction, and platelet and neutrophil activation [40]. Loss of the endothelial aryl hydrocarbon receptor (AHR) also increased tissue barrier dysfunction and subsequent movement of inflammatory cells into alveoli following influenza infection. A diet rich in AHR ligands (indoles) protected against tissue barrier dysfunction, demonstrating the importance of the gut-lung axis in viral infections [41].

Dr Kübler furthermore described novel targets for pneumonia-related acute lung injury. Cystic fibrosis (CF) transmembrane conductance regulator (CFTR), the membrane channel involved in the pathogenesis of CF, was down-regulated following *Streptococcus pneumoniae* infection. This led to endothelial barrier dysfunction through various mechanisms, including the activation of voltage-gated calcium channels and transient receptor potential vanilloid 4 (TRPV4). The CFTR potentiator ivacaftor reduced endothelial permeability following *Streptococcus pneumoniae* infection [42]. Vasculotide, agonist of the angiopoietin receptor Tie2, reduced lung permeability and acute lung injury when used with ampicillin in mice infected with *Streptococcus pneumoniae* and were mechanically ventilated [43].

This session highlighted the breadth and quality of translational respiratory research over the last year, covering many causes of impaired tissue regeneration and lung function in lung disease (Fig. 2).

Tissue modelling and remodelling – Oral presentation session

Tissue remodeling is a process occurring due to aberrant repair responses to tissue damage, leading to the loss of tissue integrity, disrupted extracellular matrix homeostasis, and replacement with disorganized structural cells [44]. Alongside fibrosis, tissue remodeling is a common feature in many respiratory diseases, such as asthma, COPD, and IPF [45]. In this session, the speakers used various experimental methodologies including murine models, human *exvivo/in vitro* cell culture models, and single cell-omics technologies to model diseased tissues and tease out the mechanisms underlying tissue remodeling.

Murine models: Mutations in surfactant protein C (SP-C) gene (*SFTPC*) in AT2 epithelial cells have been linked to sporadic and familiar IPF and a fibrotic lung phenotype [46, 47]. Using a murine model of lung fibrosis where mutant *Sftpc*^{*I*737} (I^{ER}-SP-C^{*I*73T}) was inducibly expressed, Dr Luis Rodriguez (Philadelphia, United States) showed a role for epithelial metabolic dysfunction in IPF mediated by AT2 glycolytic

reprogramming, mitochondrial dysfunction and altered AMPK signals which could be rescued by Metformin (indirect AMPK agonist). Next, Dr Sabina Janciauskiene Wallmark (Hannover, Germany) showed the beneficial effects of plasma-purified alpha1-antitrypsin therapy in preventing the development of Obliterative bronchiolitis, and attenuating acute rejection in an orthotopic model (Balb/C mice as donors and C57BL/6 as recipients) for lung transplantation [48].

Human *ex vivo/in vitro* culture models: fibroblast-derived MMPs have been postulated to be drivers of extensive lung tissue destruction and remodelling during *Mycobacterium tuberculosis* (Mtb) infection [49]. Using primary human lung fibroblasts treated with control or Mtb-infected monocytes, Miss Ramla Cusman (London, United Kingdom) showed that MMP-1 and MMP-3 were elevated in fibroblasts treated with Mtb-infected monocytes, and that inhibiting glycolysis with 2-Deoxy-D-glucose resulted in dose-dependent reduction in MMP-1 and reduction in *TIMP-1* gene expression. These results propose that fibroblast MMP and TIMP-1 secretion are monocyte-dependent and suggest host-directed strategies targeting metabolic pathways may decrease lung fibrosis in tuberculosis. Using nasal epithelial cells obtained from people with severe asthma, Dr Marianne Baastrup Soendergaard (Copenhagen, Denmark) explained that people whoare unable to down-titrate anti-IL5 tended to have impaired wound healing (determined by a wound/scratch test), suggesting that epithelial dysfunction could be a marker of incomplete remission on treatment. In this study, complete responders to anti-IL-5 had better results on lung function tests and improved symptoms compared to non-complete responders [50].

This session also included work on human induced pluripotent stem cell (hiPSC)-derived lung cells, such as a study on Birt-Hogg-Dubé syndrome (BHD), a rare autosomal dominant disorder caused by germline mutations in the tumour suppressor gene, *FLCN*, encoding for the protein folliculin [51]. Dr Alejandro Rodriguez Ruiz (Leiden, Netherlands) generated a BHD *in vitro* model by deleting *FLCN* in hiPSCs using CRISPR-Cas9 and differentiating those cells into iPSC-derived AT2 (iAT2) epithelial cells. Together with primary AT2 cells obtained from people with BHD, which were used to validate the *in vitro* model, Dr Rodriguez Ruiz utilized a lung-on-chip model to expose these cells to breathing related-stresses [52]. Additionally, hiPSCs were used by Miss Anja Schweikert (Dublin, Ireland) to generate iAT2 cells to investigate whether estradiol affects development of pulmonary fibrosis in non-diseased organoids. Epidemiological data on disease onset of IPF, as well as data in a bleomycin mouse model, suggest a role for sex hormones in disease pathogenesis [53]. Even though no significant differences were found in AT2 markers or selected proinflammatory or fibrotic genes in response to estradiol, it would be interesting to further investigate the effect of sex hormones in diseased iAT2 cells to understand potential sex-specific differences in the disease.

Single cell-omics: Mr Niklas Jonathan Lang (Munich, Germany) explained how he could observe an induction of multi-lineage conserved fibrogenic cell states by 1) coupling *ex vivo* cytokine and drug perturbations of human precision-cut lung slices (hPCLS) with scRNAseq to study early lung fibrogenesis directly in human tissue and 2) comparing the data against an *in vivo* multi-cohort single cell atlas from pulmonary fibrosis individuals. Using micro-CT staged human tissues, he characterized the appearance and interaction of *CTHRC1*⁺ myofibroblasts, *KRT17*⁺/*KRT5*⁻ basaloid epithelial cells, and an ectopic *PLVAP*⁺/*VWA1*⁺ endothelial cell state in the thickened alveolar septum of early-stage pulmonary fibrosis. This supports the use of hPCLS for drug testing and provides a framework for in-tissue perturbational single cell genomics [54]. Utilising a Multiple Iterative Labeling by Antibody

Neodeposition (MILAN) methodology on tissue sections of COPD and IPF explanted lungs, Dr Emanuela Elsa Cortesi (Leuven, Belgium) found five distinct cell clusters (basal, AT1, AT2, intermediate AT2-to-AT1, macrophages) based on 9 phenotypic markers. They also demonstrated increased levels of LGR6 in basal, AT2 cells and intermediate alveolar progenitor populations located in fibrotic regions, and in areas of inflammatory infiltration in COPD and IPF lungs that is associated with increased levels of p21 senescence marker [55]. Next, Mr Quazi Islam (Montreal, Canada) demonstrated a role for human antigen R (HuR) in lung fibroblast differentiation during IPF [56] by analysing TGF-β-treated HuR siRNA knockdown and vector-control-treated normal fibroblasts and IPF-fibroblasts using concomitant RNAseq and mass spectrometry-based proteomics techniques. Lastly, Dr Puja Mehta (London, United Kingdom) provided late-breaking data from single cell transcriptomic and T cell receptor profiles of bronchoalveolar lavage (BAL) cells obtained from people with post-COVID-19(>3 months from acute disease) who have residual lung abnormalities with predominant 1) inflammatory or 2) fibrotic radiological appearances from a CT scan. Dr Mehta showed that the two participant groups were transcriptionally similar and exhibited clonal expansion and high TCR clustering without enrichment for SARS-CoV-2 reactive sequences, indicating that the purported radiological subphenotypes in such groups may well be a different manifestation of the same disease. Therefore, T cell directed therapies might be beneficial for these people regardless of radiological appearance.

What did you always want to know about omics analyses for clinical practice? - Hot topic

Rapid advances in omics technologies have provided us with the tools to dissect biological processes at single cell resolution. Integration of omics data (multi-omics) can reveal clinically important endotypes and phenotypes, with the potential to identify new therapeutic targets.

Dr Martijn Nawijn (Groningen, the Netherlands) explained that use of transcriptomics is key to understanding how cellular activity is related to its genetic information. He focused on the transition from bulk to scRNAseq, which has revolutionised pathogenesis studies by providing in-depth analysis of differences in cell-type composition, activity and (sub)phenotype within complex samples, and information on cell-cell interactions and transitions in cell state [57]. The first study presented using scRNAseq in asthma identified a novel mucous ciliated cell state, and dominance of type 2 helper T (Th2) cell signalling [58]. Further work utilising scRNAseq showed heterogeneity within Th2 cell populations and identified a subset of pathogenic IL-9 expressing Th2 cells that was increased in allergic asthmatic individuals compared to allergic individuals without asthma [59]. Strikingly, postallergen challenge, Th2 cells were only present in the airways in asthma, and airway epithelial cells demonstrated a dramatically altered transcriptional response in subjects with asthma but not in those with allergy alone [60]. The online resource Human Lung Cell Atlas integrates multiple scRNAseq respiratory system datasets, facilitating disease comparisons at the single cell level with the potential to identify novel targets for intervention [61].

Dr Ian Adcock (London, United Kingdom) discussed the increasing sophistication of proteomic techniques, enabling selective quantification of proteins within a complex sample. Mass cytometry by time-of-flight (CyTOF) utilises heavy-metal isotope-labelled antibodies to detect and quantify multiple proteins in single cells [62]. Thus, CyTOF can identify distinct cell populations, e.g. lung adenocarcinoma-associated immune cells [63] and various immune cell populations in interstitial lung diseases [64]. Proteomic signatures can also be used to identify clinical phenotypes: sputum proteome clusters in asthma represented discrete molecular sub-phenotypes and identified candidate protein

biomarkers [65]. It was emphasized that identification of protein "hits" requires validation over time, and it remains challenging to relate cell subtype to functionality and to demonstrate disease relevance. Using machine learning, nasal fluid protein signatures were mapped to transcriptomic datasets, identifying subsets of severe asthmatics [66]. Further, differentially expressed gene/protein pathway analysis in this study revealed potential novel therapeutic targets.

Digital spatial profiling (DSP) is a complementary technique that adds a crucial layer of information, linking transcriptomics and proteomics to imaging. Dr Francesca Polverino (Houston, United States) described how spatial omics is a cutting-edge tool that allows structural navigation of the lung by digitally selecting regions of interest [67]. Identification of gene and protein enrichments within a specific spatial context using the same input material can predict pathologies associated with specific lung regions. The first DSP study in COPD demonstrated that the immune checkpoint, PD-L1, was spatially clustered with protein markers of activated T cells, as well as genes involved in cancer progression. In bronchioles, PD-L1 expression was associated with functionally active alveolar macrophages and directly correlated with lung function [68].

To fully understand heterogeneity in disease, it is vital to combine multiple omics platforms. Dr Rosa Faner Canet (Barcelona, Spain) illustrated different multi-omic integration approaches to inform clinical medicine. One approach is to use clinical phenotypes to identify underlying biological mechanisms (endotypes): COPD clusters, identified by spirometry and imaging, revealed differential protein and gene expression associated with distinct clinical outcomes [69]. Alternatively, multi-omics can expose mechanistic links that identify clinical phenotypes: integration of the sputum transcriptome, proteome and metabolome with the serum proteome demonstrated that airway microbiota metabolites may mediate COPD pathophysiology [70]. Ultimately, the approach used for multi-level integration depends on the research question, and the selection of platforms may influence the endotype uncovered.

In conclusion, this session highlighted the power of omics to reveal novel disease mechanisms and lead us towards precision medicine. Collaboration is vital, both for robustness and validation by increasing cohort size, and for multi-disciplinary interpretation of outcomes. The challenge is to integrate clinical and multi-omic data longitudinally for therapeutic translation.

Lost in translation: new insights into cell-to-cell crosstalk in lung disease – Oral presentation session

This session showcased how integration of next-generation sequencing with laboratory methods could be used to investigate cell-to-cell crosstalk (Fig. 3). First, Dr Laurens De Sadeleer (Munich, Germany) introduced epithelial-mesenchymal crosstalk. This process is vital in lung regeneration and repair after injury, and of particular interest in IPF [71, 72]. Using single-nuclei RNA sequencing (snRNAseq), laser capture microdissection, spatial transcriptomics and multiplexed immunofluorescence, novel injuryassociated profibrotic cell states were successfully identified. Importantly, further analysis revealed niche ligand-specific cell-to-cell interactions distinct between normal and early stages of IPF.

Next, Dr Nahal Mansouri (Lausanne, Switzerland) focussed on the role of basophils in regulating tumours, combining multi-level analyses of scRNAseq with complex laboratory models enabled the exploration of the role of understudied basophils and their interaction with regulatory T cells. Clinically relevant interventions (antihistamines) disturbed the interactions between basophils/Tregs,

promoting tumour progression in mice. This work highlighted a surprising role of cell-to-cell crosstalk that directly impacted the risk of metastasis in humans.

Dr Amanda Oliver (Cambridge, United Kingdom) shared a recent integrated cell atlas of healthy and diseased lungs [73]. The value of this database was demonstrated by combining scRNAseq and spatial transcriptomics to reveal novel circuits of cell communication between epithelial cells and CD4⁺T cells. They highlighted increased abundance and activation of resident memory T cells in asthmatics, an important cell type in the lung [74]. This integrated multi-omics approach identified increased interactions of goblet cells with other epithelial cells, and with CD4 T cells, which is mediated via the major histocompatibility complex in people with persistent asthma. This work provides valuable insights into targetable mechanisms behind regulatory networks of T cell activation in asthma.

Dr Jong Huat Tee (Singapore, Singapore) presented valuable insight into the anti-inflammatory role of Isthmin-1 (ISM1) in allergic asthma, whose function has been described for other conditions [75, 76]. A knockout mouse model showed that ISM1 reduces eosinophil number in BAL fluid and reduces adiponectin secretion from AT2 epithelial cells. The multicellular effect of ISM1 deficiency directly correlated with intensified inflammation, necroptosis and airway hyperresponsiveness. This presents ISM1 as a mediator of cellular interaction and a potential therapeutic tool in allergic asthma [77].

Next, Dr Dóra Paróczai (Szeged, Hungary) presented findings on extracellular neutrophil traps (NET) in airway inflammation. Charcot-Leyden crystals (CLC), known to induce neutrophil recruitment and NET formation [78], were demonstrated to have diminished effects in complement protein-depleted mice. Granulocyte-macrophage colony-stimulating factor increased uptake of CLC, increasing NET formation and complement proteins C3 and C5aR1. These findings reveal a novel therapeutic target in people with unresponsive asthma via NET-based anti-inflammatory pathways.

Dr Ken Bracke (Ghent, Belgium) utilised RNAseq to explore cellular crosstalk in COPD using B cells cocultured with fibroblasts. Together with immunohistochemical co-staining for B cells and stromal cell markers, the localisation of B cells was highlighted to impact COPD's inflammatory and remodelling pathways, building upon previous findings [79]. As such, B cells, lymphoid follicles, and fibroblasts have a dynamics role as critical regulators of COPD [80, 81]. Miss Heloisa Zimermam (Ribeirao Preto, Brazil) explored the complex immune cell communication networks. Dendritic cells have a protective role in tumour microenvironments [82, 83]. The findings of this work show ineffective dendritic cells function in the tumoral front area. This was specific to adenocarcinoma as opposed to squamous cell carcinoma. Therefore, cell interactions are disease subtype-specific, informing therapeutic interventions.

Returning to COPD, Dr Harriet Owles (London, United Kingdom) focussed on IL36 γ and its effects on lung macrophages [84]. Supernatants from IL36 γ -stimulated small airway fibroblasts were exposed to monocyte-derived macrophages. Here, they show that increased levels of IL36 γ impair macrophage phagocytosis in COPD. Notably, IL36 γ expression/release is increased by viral infection [85], making this novel cell-cell crosstalk relevant for acute COPD exacerbations [86].

Finally, Dr Caroline Lindo (Lund, Sweden) used surgical lung tissue samples to reveal the relationship between eosinophils, microbes, and immune cell patterns [87]. Combining *in situ* hybridisation, multiplexed immunohistochemistry and spatial analysis enabled an investigation of immune infiltration patterns. No spatial correlation of infiltration with bacteria, viruses or fungi was found. However, spatially distinct cell niches were revealed. Eosinophils and a type 2 inflammatory features

were linked with basophils, indicating a spatial correlation [85]. This patchy pattern of immune cell niches results in a complex mix of inflammatory signatures, which impacts treatment effectiveness [88].

From the transcriptome landscape to innovative preclinical models in lung diseases – Oral presentation session

This session highlighted a variety of state-of-the-art, innovative approaches to explore complex aspects of lung diseases, providing valuable insights into chronic airway diseases, pulmonary fibrosis, post-infection complications, and conditions like COPD that result in skeletal muscle wasting.

Dr Arnaud Bourdin (Montpellier, France) presented a novel model combining the bronchial epithelium and submucosa, both playing an important role in many chronic airway diseases including asthma. The model consists of human bronchial fibroblasts seeded on a collagen-chitosan matrix, iPSC-derived bronchial epithelial cells (forming basal, goblet, club and neuroendocrine cells) [89] and iPSC-derived neurons. To facilitate axonal integration into the existing airway epithelium, Schwann cells were added (previously described to improve nerve regeneration [90]), resulting in an improved innervation of the airway epithelium that can be used to model chronic airway diseases.

Ultra-strong exercise induces physiological responses in the human body. Dr Agnieszeka Smolinska (Maastricht, the Netherlands) described that volatile organic compounds (VOCs), which can be measured in exhaled breath using high-resolution TD-GC-MS, change after running an ultra-marathon. Breath was collected pre- and post-ultra-marathon from 24 healthy participants. Here, 811 VOCs were differentially regulated, with 12 being significantly decreased and 51 significantly increased post-ultra-marathon. Seven of the significantly upregulated compounds after the ultra-marathon suggest physiological responses like fatty acid oxidation, inflammation and altered gut microbiome activity.

Lung explants mostly recapitulate the end stage of IPF, limiting the outcome of these models. Additionally, scRNAseq studies of lung explants are lacking spatial information [91-94]. Dr Aurélien Justet (Caen, France) applied high-resolution spatial transcriptomics to earliest clinical-grade IPF samples to recapitulate the architecture of the human airways. Early disease was characterized by change in cell type proportions (decreased AT1, AT2 cells and general capillaries) and increased COL15 venous and ectopic airway cells, suggesting respiratory unit loss. Thus, spatial transcriptomic analysis allows the investigation of cellular changes of the alveolar niches.

Dr Ana Lilia Serna Valverde (Nottingham, United Kingdom) used a hiPSC-derived model with a *SFTPC* mutation generated from an individual with IPF and CRISPR gene-edited wild-type control [95, 96] to investigate the impact of the *SFTPC* mutation on the iAT2 cell response to infection with Influenza A virus subtype H1N1 [97, 98]. Interestingly, bulk RNAseq revealed, in addition to top genes involved in IPF and infection, that wild-type cells mainly show GO terms associated to the anti-bacterial defence response, whilst the mutant cell line mainly displayed GO terms associated with the cells' reaction to its environment. This model demonstrates the potential of gene-edited iAT2s as *in vitro* platforms for human respiratory infection modelling.

Dr Yuki Yamamoto (Kyoto, Japan) presented an IPF model composed of healthy iPSC-derived alveolar organoids [99] co-cultured with lung fibroblasts. scRNAseq showed that this model after treatment with bleomycin recapitulated key mechanisms of fibrosis sharing 76.3% of upregulated pathways with

IPF human-derived lung samples. Treating these fibrotic organoids with HL001, a LPA1 antagonist [100], showed a restorative effect with decrease of fibrosis, and increase of AT2 cell marker. Consistent with a previous report [101], murine and human organoid models proved the effectiveness of HL001 in IPF.

By combining hPCLS generated from lung tissue from IPF donors with snRNAseq Dr Martin Decaris (San Francisco, United States) investigated bexotegrast, a dual $\alpha V\beta 6/\alpha V\beta 1$ integrin inhibitor in the fibrotic lung models. He showed that bexotegrast reduces ECM-related gene expression in fibroblasts, attenuates CTHRC1⁺ pro-fibrotic fibroblast subpopulation and reduces fibrogenic gene expression pathways in aberrant basaloid cells [102].

Lung tissue biopsies may aid the diagnosis of fibrotic interstitial lung diseases (ILDs); however, less invasive alternatives are needed. Dr Avraham Unterman (Tel Aviv, Israel) used scRNAseq to investigate novel biomarkers in BAL [103, 104] by characterizing differences in BAL composition between fibrotic hypersensitivity pneumonitis (fHP) and IPF. They found that the proportions of non-FABP4⁺ macrophages, regulatory T cells and CLEC9A⁺ dendritic cells are significantly increased in fHP vs IPF. In addition, fHP macrophages showed a pro-inflammatory activation pattern. These findings may help to differentiate IPF from fHP without the need of invasive techniques.

Mr Mohammad Shadab Ali (New Delhi, India), delved into the molecular underpinnings of the severe post-infection complication known as post-COVID pulmonary fibrosis (PCPF) [105]. This was achieved by comparing BAL samples obtained from people with PCPF with those obtained from non-ILD individuals. Analyses of KEGG and IPA pathways unveiled the involvement of pathways associated with the nervous system in PCPF, and identified key regulators play a crucial role in the cytoskeleton organization. The insights gained from this molecular investigation enhance our comprehension of PCPF and present potential therapeutic targets.

Next, Dr Pauline Henrot (Pessac, France) explored the involvement of CXCR4⁺ cells [106] in skeletal muscle wasting among people with COPD [107]. Using an early COPD mouse model with a CXCR4 deletion, the study found that this deletion prevented a decrease in muscle endurance and the loss of oxidative myofibers. Dr Henrot intends to employ snRNAseq to further analyze the inflammatory infiltrate and dysregulated pathways.

Collectively, this session showcased innovative approaches in utilising transcriptomics (Table 1) to advance our understanding of disease mechanisms and identify potential drug targets. This emphasizes the significance of continued research in this field.

Concluding remarks

The selected sessions summarised in this review article showcased the diversity in basic and translational respiratory science and the remarkable progress presented at this year ERS congress. The studies delved into the intricate interplay of micro- and macro-environmental factors impacting respiratory health, emphasizing the urgency for comprehensive strategies addressing both environmental influences and individual behaviours. They illuminated the transformative potential of omics technologies, revealing cellular states and interactions that were previously unseen and paving the way for precision medicine. The exploration of cell-to-cell crosstalk provided deep insights into the complex networks underlying lung diseases, offering promising avenues for targeted interventions.

Additionally, innovative preclinical models and advanced molecular analyses unveiled novel aspects of various lung conditions, laying the groundwork for future research and therapeutic development. The topics discussed at the ERS congress 2023 collectively underscored the collaborative efforts and interdisciplinary approaches driving the advancements in respiratory science, offering hope for improved treatments and a healthier respiratory future for people worldwide.

Table 1: Summary of the presented innovative approaches to model lung diseases presented as part of the 'From the transcriptome landscape to innovative preclinical models in lung diseases' oral presentation session.

Models	Transbronchial	hPCLS	BAL	Exhaled	iPSC-derived	Mouse
used	cryobiopsy			breath	models	model
Diseases/co	IPF	IPF	IPF, fHP, hPCLS	Ultra-strong	Asthma, IPF	COPD
nditions				exercise		
investigated						
Purpose	Investigate	Test	Investigate	Investigate	Create novel	Investigate
	cellular changes	drug	biomarkers to	physiological	innervated	role of
	of the alveolar	bexote	characterize	response	airway	CXCR4 in
	niche in IPF	grast	differences	[112]	epithelium	skeletal
	[108]	[109]	between fHP		model [113],	muscle
			and IPF [110],		model	wasting
			Investigate		respiratory	[115]
			molecular basis		infection with	
			of PCPF [111]		H1N1 [98], test	
					drug HL001	
					[114]	
Read-out	High resolution	snRNAs	scRNAseq	High	Bulk RNAseq	Functional
	spatial	eq		resolution	and scRNAseq	tests and
	transcriptomics			TD-GC-MS		whole
						tissue
						proteomics

References

1. Ocaña SC, El-Merhie N, Kuipers ME, et al. ERS international Congress 2022: highlights from the Basic and Translational Science Assembly. *ERJ Open Research* 2023: 00561-02022.

2. Ubags ND, Baker J, Boots A, et al. ERS International Congress, Madrid, 2019: highlights from the Basic and Translational Science Assembly. *ERJ Open Res* 2020: 6(1).

3. Nikolić MZ, Garrido-Martin EM, Greiffo FR, et al. From the pathophysiology of the human lung alveolus to epigenetic editing: Congress 2018 highlights from ERS Assembly 3 "Basic and Translational Science.". *ERJ Open Res* 2019: 5(2).

4. Farr A, Ocaña SC, Gille T, et al. What to expect from the ERS International Congress 2023. *Breathe* 2023: 19(2): 230107.

5. Son K, Landt EM, Fisser C, et al. Interview with the ECM Award winner 2022 and introducing the new ECM members. *Breathe (Sheff)* 2023: 19(1): 220274.

6. Backman H, Blomberg A, Lundquist A, et al. Lung Function Trajectories and Associated Mortality Among Adults with and without Airway Obstruction. *Am J Respir Crit Care Med* 2023. 7. Lindberg A, Lundbäck B. The Obstructive Lung Disease in Northern Sweden Chronic Obstructive Pulmonary Disease Study: design, the first year participation and mortality. *Clin Respir J* 2008: 2 Suppl 1: 64-71.

8. Livestock farming and residential health. [cited 01.10.2023]; Available from: https://www.rivm.nl/en/livestock-farming-and-health/livestock-farming-and-residential-health

 van Kersen W, Bossers A, de Steenhuijsen Piters WAA, et al. Air pollution from livestock farms and the oropharyngeal microbiome of COPD patients and controls. *Environ Int* 2022: 169: 107497.
 Lanza VF, Baquero F, Martínez JL, et al. In-depth resistome analysis by targeted

metagenomics. Microbiome 2018: 6(1): 11.

11. Borlée F, Yzermans CJ, van Dijk CE, et al. Increased respiratory symptoms in COPD patients living in the vicinity of livestock farms. *Eur Respir J* 2015: 46(6): 1605-1614.

12. Amin H, Cramer C, Drengenes C, et al. Association between indoor bacterial communities, lung function and airway inflammation. *European Respiratory Journal* 2023: 62(suppl 67): OA2517.

13. Yu Z, Ekström S, Bellander T, et al. Ambient air pollution exposure linked to long COVID among young adults: a nested survey in a population-based cohort in Sweden. *Lancet Reg Health Eur* 2023: 28: 100608.

14. Yu Z, Bellander T, Bergström A, et al. Association of Short-term Air Pollution Exposure With SARS-CoV-2 Infection Among Young Adults in Sweden. *JAMA Netw Open* 2022: 5(4): e228109.

15. Björkander S, Du L, Zuo F, et al. SARS-CoV-2-specific B- and T-cell immunity in a populationbased study of young Swedish adults. *J Allergy Clin Immunol* 2022: 149(1): 65-75.e68.

16. Valencia-Hernandez C, Gehring U, Koppelman GH, et al. The residential green environment and lung function into adolescence in European birth cohorts. A CADSET initiative. *European Respiratory Journal* 2023: 62(suppl 67): OA2519.

17. Paciência I, Rantala AK, Antikainen H, et al. Varying effects of greenness in the spring and summer on the development of allergic rhinitis up to 27 years of age: The Espoo Cohort Study. *Allergy* 2023: 78(6): 1680-1682.

18. So R, Lim Y-H, Zhang J, et al. Long-term exposure to air pollution and the risk of obstructive lung disease in a Danish nationwide analysis. *European Respiratory Journal* 2023: 62(suppl 67): OA2521.

19. Schmidt M, Schmidt SA, Sandegaard JL, et al. The Danish National Patient Registry: a review of content, data quality, and research potential. *Clin Epidemiol* 2015: 7: 449-490.

20. Pottegård A, Schmidt SAJ, Wallach-Kildemoes H, et al. Data Resource Profile: The Danish National Prescription Registry. *Int J Epidemiol* 2017: 46(3): 798-798f.

21. de Hoogh K, Chen J, Gulliver J, et al. Spatial PM2.5, NO2, O3 and BC models for Western Europe – Evaluation of spatiotemporal stability. *Environment International* 2018: 120: 81-92.

22. Zhao T, Markevych I, Fuertes E, et al. Impact of long-term exposure to ambient ozone on lung function over a course of 20 years (The ECRHS study): a prospective cohort study in adults. *Lancet Reg Health Eur* 2023: 34: 100729.

23. Markevych I, Zhao T, Fuertes E, et al. Residential greenspace and lung function decline over 20 years in a prospective cohort: The ECRHS study. *Environment international* 2023: 178: 108036.

 Kothe H, Cuesta A, Madueno L, et al. Influence of improved cookstove in rural Rwanda on indoor air pollution and lung function. *European Respiratory Journal* 2023: 62(suppl 67): OA2523.
 Soriano JB, Solozabal-Coll M, Galindo JV, et al. Economic and environmental impact of the

non-clinical change of inhaler devices for COPD and asthma in Spain. *European Respiratory Journal* 2023: 62(suppl 67): OA2524.

26. Pulakka A, Risnes K, Metsälä J, et al. Preterm birth and asthma and COPD in adulthood: a nationwide register study from two Nordic countries. *Eur Respir J* 2023: 61(6).

27. Nissen G, Hinsenbrock S, Rausch TK, et al. Lung Function of Preterm Children Parsed by a Polygenic Risk Score for Adult COPD. *NEJM Evidence* 2023: 2(3): EVIDoa2200279.

28. Dapas M, Thompson EE, Wentworth-Sheilds W, et al. Multi-omic association study identifies DNA methylation-mediated genotype and smoking exposure effects on lung function in children living in urban settings. *PLoS Genet* 2023: 19(1): e1010594.

29. Wang T, Jia Z, Li S, et al. The association between leukocyte telomere length and chronic obstructive pulmonary disease is partially mediated by inflammation: a meta-analysis and population-based mediation study. *BMC Pulm Med* 2022: 22(1): 320.

30. Jeridi A, Günsel GG, Novikava M, et al. Ferroptosis, induced by macrophages drives COPD pathogenesis. *ERJ Open Research* 2022: 8(suppl 8): 177.

31. Liang J, Huang G, Liu X, et al. The ZIP8/SIRT1 axis regulates alveolar progenitor cell renewal in aging and idiopathic pulmonary fibrosis. *J Clin Invest* 2022: 132(11).

32. Moimas S, Salton F, Kosmider B, et al. miR-200 family members reduce senescence and restore idiopathic pulmonary fibrosis type II alveolar epithelial cell transdifferentiation. *ERJ Open Res* 2019: 5(4).

33. Volpe MC, Ciucci G, Zandomenego G, et al. Flt1 produced by lung endothelial cells impairs ATII cell transdifferentiation and repair in pulmonary fibrosis. *Cell Death Dis* 2023: 14(7): 437.

34. Rustam S, Hu Y, Mahjour SB, et al. A Unique Cellular Organization of Human Distal Airways and Its Disarray in Chronic Obstructive Pulmonary Disease. *Am J Respir Crit Care Med* 2023: 207(9): 1171-1182.

35. Jaeger B, Schupp JC, Plappert L, et al. Airway basal cells show a dedifferentiated KRT17(high)Phenotype and promote fibrosis in idiopathic pulmonary fibrosis. *Nat Commun* 2022: 13(1): 5637.

36. Ahangari F, Becker C, Foster DG, et al. Saracatinib, a Selective Src Kinase Inhibitor, Blocks Fibrotic Responses in Preclinical Models of Pulmonary Fibrosis. *Am J Respir Crit Care Med* 2022: 206(12): 1463-1479.

37. ClinicalTrials.gov. [cited; Available from: <u>https://clinicaltrials.gov/study/NCT01915511</u>

38. Caporarello N, Lee J, Pham TX, et al. Dysfunctional ERG signaling drives pulmonary vascular aging and persistent fibrosis. *Nat Commun* 2022: 13(1): 4170.

39. Bian F, Lan YW, Zhao S, et al. Lung endothelial cells regulate pulmonary fibrosis through FOXF1/R-Ras signaling. *Nat Commun* 2023: 14(1): 2560.

40. Jandl K, Berg JL, Birnhuber A, et al. Basement membrane product, endostatin, as a link between inflammation, coagulation and vascular permeability in COVID-19 and non-COVID-19 acute respiratory distress syndrome. *Front Immunol* 2023: 14: 1188079.

41. Major J, Crotta S, Finsterbusch K, et al. Endothelial AHR activity prevents lung barrier disruption in viral infection. *Nature* 2023: 621(7980): 813-820.

42. Erfinanda L, Zou L, Gutbier B, et al. Loss of endothelial CFTR drives barrier failure and edema formation in lung infection and can be targeted by CFTR potentiation. *Sci Transl Med* 2022: 14(674): eabg8577.

43. Lask A, Gutbier B, Kershaw O, et al. Adjunctive therapy with the Tie2 agonist Vasculotide reduces pulmonary permeability in Streptococcus pneumoniae infected and mechanically ventilated mice. *Sci Rep* 2022: 12(1): 15531.

44. Liu G, Philp AM, Corte T, et al. Therapeutic targets in lung tissue remodelling and fibrosis. *Pharmacol Ther* 2021: 225: 107839.

45. Singla A, Reuter S, Taube C, et al. The molecular mechanisms of remodeling in asthma, COPD and IPF with a special emphasis on the complex role of Wnt5A. *Inflamm Res* 2023: 72(3): 577-588.

46. Cameron HS, Somaschini M, Carrera P, et al. A common mutation in the surfactant protein C gene associated with lung disease. *J Pediatr* 2005: 146(3): 370-375.

47. Nureki SI, Tomer Y, Venosa A, et al. Expression of mutant Sftpc in murine alveolar epithelia drives spontaneous lung fibrosis. *J Clin Invest* 2018: 128(9): 4008-4024.

48. Nakagiri T, Wrenger S, Sivaraman K, et al. α1-Antitrypsin attenuates acute rejection of orthotopic murine lung allografts. *Respir Res* 2021: 22(1): 295.

49. Elkington PT, Ugarte-Gil CA, Friedland JS. Matrix metalloproteinases in tuberculosis. *Eur Respir J* 2011: 38(2): 456-464.

50. Soendergaard MB, Hansen S, Bjerrum AS, et al. Complete response to anti-interleukin-5 biologics in a real-life setting: results from the nationwide Danish Severe Asthma Register. *ERJ Open Res* 2022: 8(4).

51. Daccord C, Good JM, Morren MA, et al. Birt-Hogg-Dubé syndrome. *Eur Respir Rev* 2020: 29(157).

52. van Riet S, van Schadewijk A, Khedoe P, et al. Organoid-based expansion of patient-derived primary alveolar type 2 cells for establishment of alveolus epithelial Lung-Chip cultures. *Am J Physiol Lung Cell Mol Physiol* 2022: 322(4): L526-I538.

53. Zaman T, Lee JS. Risk factors for the development of idiopathic pulmonary fibrosis: A review. *Curr Pulmonol Rep* 2018: 7(4): 118-125.

54. Lam M, Lamanna E, Organ L, et al. Perspectives on precision cut lung slices-powerful tools for investigation of mechanisms and therapeutic targets in lung diseases. *Front Pharmacol* 2023: 14: 1162889.

55. Cortesi EE, Meeusen B, Vanstapel A, et al. Increased LGR6 Expression Sustains Long-Term Wnt Activation and Acquisition of Senescence in Epithelial Progenitors in Chronic Lung Diseases. *Cells* 2021: 10(12).

56. Trivlidis J, Aloufi N, Al-Habeeb F, et al. HuR drives lung fibroblast differentiation but not metabolic reprogramming in response to TGF-β and hypoxia. *Respir Res* 2021: 22(1): 323.

57. Alexander MJ, Budinger GRS, Reyfman PA. Breathing fresh air into respiratory research with single-cell RNA sequencing. *Eur Respir Rev* 2020: 29(156).

58. Vieira Braga FA, Kar G, Berg M, et al. A cellular census of human lungs identifies novel cell states in health and in asthma. *Nat Med* 2019: 25(7): 1153-1163.

59. Seumois G, Ramirez-Suastegui C, Schmiedel BJ, et al. Single-cell transcriptomic analysis of allergen-specific T cells in allergy and asthma. *Sci Immunol* 2020: 5(48).

60. Alladina J, Smith NP, Kooistra T, et al. A human model of asthma exacerbation reveals transcriptional programs and cell circuits specific to allergic asthma. *Sci Immunol* 2023: 8(83): eabq6352.

61. Sikkema L, Ramirez-Suastegui C, Strobl DC, et al. An integrated cell atlas of the lung in health and disease. *Nat Med* 2023: 29(6): 1563-1577.

62. Iyer A, Hamers AAJ, Pillai AB. CyTOF((R)) for the Masses. *Front Immunol* 2022: 13: 815828.
63. Lavin Y, Kobayashi S, Leader A, et al. Innate Immune Landscape in Early Lung

Adenocarcinoma by Paired Single-Cell Analyses. *Cell* 2017: 169(4): 750-765 e717.

64. Hata K, Yanagihara T, Matsubara K, et al. Mass cytometry identifies characteristic immune cell subsets in bronchoalveolar lavage fluid from interstitial lung diseases. *Front Immunol* 2023: 14: 1145814.

65. Schofield JPR, Burg D, Nicholas B, et al. Stratification of asthma phenotypes by airway proteomic signatures. *J Allergy Clin Immunol* 2019: 144(1): 70-82.

66. Agache I, Shamji MH, Kermani NZ, et al. Multidimensional endotyping using nasal proteomics predicts molecular phenotypes in the asthmatic airways. *J Allergy Clin Immunol* 2023: 151(1): 128-137.

67. Yang CX, Tomchaney M, Landecho MF, et al. Lung Spatial Profiling Reveals a T Cell Signature in COPD Patients with Fatal SARS-CoV-2 Infection. *Cells* 2022: 11(12).

68. Polverino F, Mirra D, Yang CX, et al. Similar programmed death ligand 1 (PD-L1) expression profile in patients with mild COPD and lung cancer. *Sci Rep* 2022: 12(1): 22402.

69. Gregory A, Xu Z, Pratte K, et al. Clustering-based COPD subtypes have distinct longitudinal outcomes and multi-omics biomarkers. *BMJ Open Respir Res* 2022: 9(1).

70. Yan Z, Chen B, Yang Y, et al. Multi-omics analyses of airway host-microbe interactions in chronic obstructive pulmonary disease identify potential therapeutic interventions. *Nat Microbiol* 2022: 7(9): 1361-1375.

71. Bagnato G, Harari S. Cellular interactions in the pathogenesis of interstitial lung diseases. *Eur Respir Rev* 2015: 24(135): 102-114.

72. Zhou H, Fan EK, Fan J. Cell-Cell Interaction Mechanisms in Acute Lung Injury. *Shock* 2021: 55(2): 167-176.

73. Kathiriya JJ, Wang C, Zhou M, et al. Human alveolar type 2 epithelium transdifferentiates into metaplastic KRT5(+) basal cells. *Nat Cell Biol* 2022: 24(1): 10-23.

74. Marone G, Schroeder JT, Mattei F, et al. Is There a Role for Basophils in Cancer? *Front Immunol* 2020: 11: 2103.

75. Sikkema L, Ramírez-Suástegui C, Strobl DC, et al. An integrated cell atlas of the lung in health and disease. *Nat Med* 2023: 29(6): 1563-1577.

76. Purwar R, Campbell J, Murphy G, et al. Resident Memory T Cells (TRM) Are Abundant in Human Lung: Diversity, Function, and Antigen Specificity. *PLOS ONE* 2011: 6(1): e16245.

77. Nguyen N, Xu S, Lam TYW, et al. ISM1 suppresses LPS-induced acute lung injury and postinjury lung fibrosis in mice. *Molecular Medicine* 2022: 28(1): 72.

78. Rivera-Torruco G, Martínez-Mendiola CA, Angeles-Floriano T, et al. Isthmin 1 is Expressed by Progenitor-Like Cells in the Lung: Phenotypical Analysis of Isthmin 1(+) Hematopoietic Stem-Like Cells in Homeostasis and during Infection. *J Immunol Res* 2022: 2022: 2909487.

79. Lam TYW, Nguyen N, Peh HY, et al. ISM1 protects lung homeostasis via cell-surface GRP78mediated alveolar macrophage apoptosis. *Proc Natl Acad Sci U S A* 2022: 119(4).

 Gevaert E, Delemarre T, De Volder J, et al. Charcot-Leyden crystals promote neutrophilic inflammation in patients with nasal polyposis. *J Allergy Clin Immunol* 2020: 145(1): 427-430.e424.
 Brusselle GG, Demoor T, Bracke KR, et al. Lymphoid follicles in (very) severe COPD: beneficial

or harmful? *Eur Respir J* 2009: 34(1): 219-230. 82. Caramori G, Casolari P, Barczyk A, et al. COPD immunopathology. *Semin Immunopathol* 2016: 38(4): 497-515.

83. Chung KF, Adcock IM. Multifaceted mechanisms in COPD: inflammation, immunity, and tissue repair and destruction. *Eur Respir J* 2008: 31(6): 1334-1356.

84. Gupta YH, Khanom A, Acton SE. Control of Dendritic Cell Function Within the Tumour Microenvironment. *Front Immunol* 2022: 13: 733800.

85. Baker JR, Fenwick PS, Koss CK, et al. IL-36 receptor agonist and antagonist imbalance drives neutrophilic inflammation in COPD. *JCI Insight* 2022: 7(15).

86. Ma Y, Shurin GV, Peiyuan Z, et al. Dendritic cells in the cancer microenvironment. *J Cancer* 2013: 4(1): 36-44.

87. Kovach MA, Singer B, Martinez-Colon G, et al. IL-36γ is a crucial proximal component of protective type-1-mediated lung mucosal immunity in Gram-positive and -negative bacterial pneumonia. *Mucosal Immunol* 2017: 10(5): 1320-1334.

88. Stavropoulou E, Kantartzi K, Tsigalou C, et al. Unraveling the Interconnection Patterns Across Lung Microbiome, Respiratory Diseases, and COVID-19. *Front Cell Infect Microbiol* 2020: 10: 619075.

89. Ahmed E FM, Bourguignon C, Mianné, Petit A, Jory M, Cazevieille C, Boukhaddaoui H, Garnett JP, Hirtz C, Massiera G, Vachier I, Assou S, Bourdin A, De Vos J. Differentiation of Human Induced Pluripotent Stem Cells from Patients with Severe COPD into Functional Airway Epithelium *Cells* 2022: 11.

90. Blais M GM, Berthod F. Improvement of Nerve Regeneration in Tissue-Engineered Skin Enriched with Schwann Cells. *Soc Inv Derm* 2009: 129.

91. Travaglini KJ NA, Penland L, Sinha R, Gillich A, Sit RV, Chang S, Conley SD, Mori Y, Seita J, Berry GJ, Shrager JB, Metzger RJ, Kuo CS, Neff N, Weissman IL, Quake SR, Krasnow MA. A molecular cell atlas of the human lung from single-cell RNA sequencing. *Nature* 2020: 587.

92. Tsukui T SK, Wetter JB, Wilson-Kanamori JR, Hazelwood LA, Henderson NC, Adams TS, Schupp JC, Poli SD, Rosas IO, Kaminski N, Matthay MA, Wolters PJ, Sheppard D. Collagen-producing lung cell atlas identifies multiple subsets with distinct localization and relevance to fibrosis. *Nat Com* 2020: 11.

93. Adams TS SJ, Poli S, Ayaub EA, Neumark N, Ahangari F, Chu SG, Raby BA, Deluliis G, Januszyk M, Duan Q, Arnett HA, Siddiqui A, Washko GR, Homer R, Yan X, Rosas IO, Kaminski N. Single-cell RNAseq reveals ectopic and aberrant lung-resident cell populations in idiopathic pulmonary fibrosis *Science Advances* 2020: 6.

94. Habermann AC GA, Bui LT, Yahn SL, Winters NI, Calvi CL, Peter L, Chung MI, Taylor CJ, Jetter C, Raju L, Roberson J, Ding G, Wood L, Sucre JMS, Richmond BW, Serezani AP, McDonnell WJ, Mallal SB, Bacchetta MJ, Loyd JE, Shaver CM, Ware LB, Bremner R, Walia R, Blackwell TS, Banovich NE, Kropsky JA. Single-cell RNA sequencing reveals profibrotic roles of distinct epithelial and mesenchymal lineages in pulmonary fibrosis. *science advances* 2020: 6.

95. Cuevas-Ocaña S YJ, Aushev M, Schlossmacher G, Bear CE, Hannan NRF, Perkins ND, Rossant J, Wong AP, Gray MA. A Cell-Based Optimised Approach for Rapid and Efficient Gene Editing of Human Pluripotent Stem Cells. *Int J Mol Sci* 2023: 24.

96. Ocaña SC, Valverde ALS, Reed L, et al. Next generation of gene-editing technologies for respiratory research and medicine. *European Respiratory Journal* 2022: 60(suppl 66): 2815.

97. Valverde ALS, Reed L, Ocaña SC, et al. In vitro modelling of respiratory infections in Idiopathic Pulmonary Fibrosis using patient-specific hiPSC-derived alveolar epithelial cells. *ERJ Open Research* 2023: 9(suppl 10): 79.

98. Valverde ALS, Reed L, Ocaña SC, et al. LSC - 2023 - In vitro modelling of respiratory infections in Idiopathic Pulmonary Fibrosis using patient-specific hiPSC-derived alveolar epithelial cells. *European Respiratory Journal* 2023: 62(suppl 67): OA897.

99. Yamamoto Y GS, Korogi Y, Seki M, Konoshi S, Ikeo S, Sone N, Nagasaki T, Matsumoto H, Muro S, Ito I, Hirai T, Kohno T, Suzuki Y, Mishima M. Long-term expansion of alveolar stem cells derived from human iPS cells in organoids. *Nature Methods* 2017: 14.

100. Tager AM LP, Shea BS, Campanella GS, Selman M, Zhao Z, Polosukhin V, Wain J, Karimi-Shah BA, Kim ND, Hart WK, Pardo A, Blackwell TS, Xu Y, Chun J, Luster AD. The lysophosphatidic acid receptor LPA1 links pulmonary fibrosis to lung injury by mediating fibroblast recruitment and vascular leak *Nature Medicine* 2008: 14.

101. Funke M ZZ, Xu Y, Chun J, Tager AM. The Lysophosphatidic Acid Receptor LPA1 Promotes Epithelial Cell Apoptosis after Lung Injury *Am J Respir Cell Mol Biol* 2012: 46.

102. Adams TS, Schupp JC, Poli S, et al. Single-cell RNA-seq reveals ectopic and aberrant lungresident cell populations in idiopathic pulmonary fibrosis. *Sci Adv* 2020: 6(28): eaba1983.

103. Raghu G R-JM, Ryerson CJ, Myers JL, Kreuter M, Vasakova M, Bargagli E, Chung JH,, Collins BF, Bendstrup E, Chami HA, Chua AT, Corte TJ, Dalphin JC, DAnoff SK, Diaz-Mendoza J, Duggal A, Egashira R, Ewing T, Gulati M, Inoue Y, Jenkins AR, Johannson KA, Johkoh T, Tamae-Kakazu M, Kitaichi M, Knight SL, Koschel D, Lederer DJ, Mageto Y, Maier LA, Matiz C, Morell F, Nicholson AG, Patolia S, Pereira CA, Renzoni EA, Salisbury ML, Selman M, Wash SLF, Wuyts WA, Wilson KC. Diagnosis of Hypersensitivity Pneumonitis in Adults. An Official ATS/JRS/ALAT Clinical Practice Guideline. *Am J Respir Crit Care Med* 2020: 202.

104. Freund O HY, Shalmon T, Wand O, Schneer S, Perluk TM, Kleinhendler E, Hershko T, Tiran B, Aviram G, Gershman E, Adir Y, Shitrit D, Bar-Shai A, Unterman A. Real-Life Diagnostic Performance of the Hypersensitivity Pneumonitis Guidelines: A Multicenter Cohort Study. *Diagnostics* 2023: 13. 105. Spagnolo P BE, Aliberti S, Cocconcelli E, Biondini D, Casa GD, Sverzellati N, Maher TM.

Pulmonary fibrosis secondary to COVID-19: a call to arms? Lancet 2020: 8.

106. Strieter RM GB, Keane MP. The role of CXC chemokines in pulmonary fibrosis. *J Clin Invest* 2007: 117: 549-556.

107. Henrot P DI, Schilfarth P, Esteves P, Blervaque L, Zysman M, Gouzi F, Hayot M, Pomiès P, Berger P. Main Pathogenic Mechanisms and Recent Advances in COPD Peripheral Skeletal Muscle Wasting. *Int J Mol Sci* 2023: 24.

108. Justet A, Zhao J, Adams T, et al. Identification of abnormal airway niches in the fibrotic lung using Spatial Transcriptomics. *European Respiratory Journal* 2023: 62(suppl 67): OA894.

109. Decaris M, An M, Ahn R, et al. Dual $\alpha V\beta 6/\alpha V\beta 1$ integrin inhibitor bexotegrast reduces fibrogenesis in pathological cell populations present in the fibrotic human lung. *European Respiratory Journal* 2023: 62(suppl 67): OA899.

110. Unterman A, Bar-Lev TH, Stein Y. Single-cell RNA sequencing of bronchoalveolar lavage reveals pro-inflammatory macrophage activation in fibrotic hypersensitivity pneumonitis. *European Respiratory Journal* 2023: 62(suppl 67): OA895.

111. Ali MS, Hadda V, Chopra A, et al. Transcriptomic landscape of post-covid-19 pulmonary fibrosis (PCPF): insights and implications of bronchoalveolar lavage (BAL). *European Respiratory Journal* 2023: 62(suppl 67): OA896.

112. Chou H, Arthur K, Shaw E, et al. Late Breaking Abstract - Volatile organic compounds in exhaled breath reflect physiological changes in ultramarathon runners. *European Respiratory Journal* 2023: 62(suppl 67): OA893.

113. FOISSET F, Lehalle C, Nasri A, et al. Late Breaking Abstract - Development of a bronchial epithelium with a sensory innervation both derived from induced pluripotent stem cells. *European Respiratory Journal* 2023: 62(suppl 67): OA892.

114. Yamamoto Y, Nagamoto T, Choi H, et al. Preclinical evaluation of HL001, a novel lysophosphatidic acid 1 (LPA1) receptor antagonist, for idiopathic pulmonary fibrosis utilizing physiologically relevant stem cell-derived lung organoids. *European Respiratory Journal* 2023: 62(suppl 67): OA898.

115. Henrot P, Schilfarth P, Campagnac M, et al. Involvement of the CXCR4/CXCL12 axis in skeletal muscle wasting in a murine model of Chronic Obstructive Pulmonary Disease (COPD). *European Respiratory Journal* 2023: 62(suppl 67): OA900.

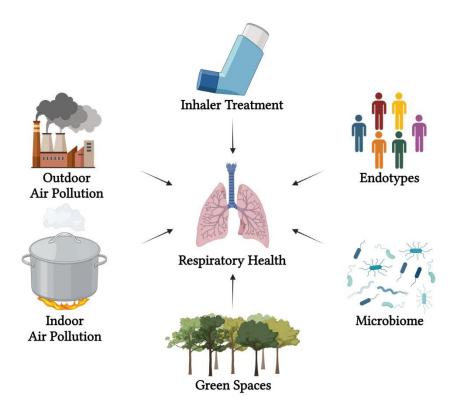


Figure 1. Schematic of the discussed environmental factors that affect lung function, including the microenvironment level of pathophysiological changes (endotypes), and microbiome, the macroenvironment level of indoor and outdoor air pollution and green spaces, as well as the use of inhaler treatment. Created with BioRender.com.

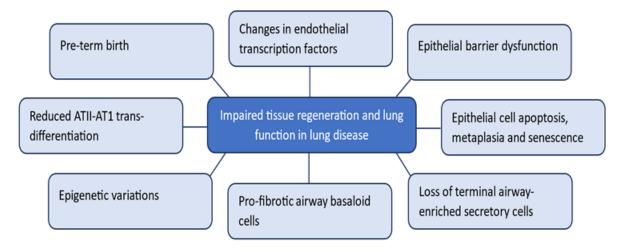


Figure 2. Causes of impaired tissue regeneration and lung function in lung disease.

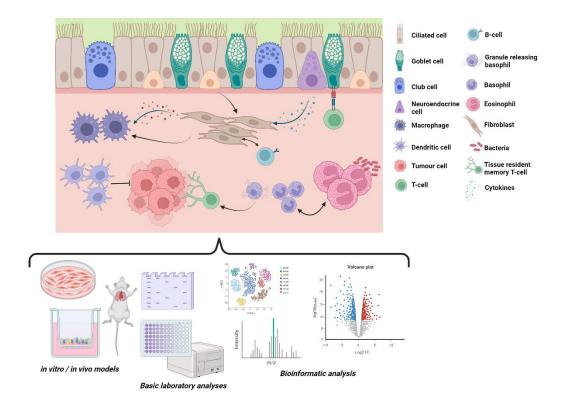


Figure 3. Cell crosstalk in the airways is highly complex, thus, there is a need to integrate basic laboratory techniques, multi-omics and bioinformatic analyses to holistically understand these interactions. Created with BioRender.com.